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Seed amplification assay results illustrate discrepancy in Parkinson's disease clinical diagnostic accuracy and error rates

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Abstract

Parkinson's disease (PD) may be misdiagnosed due to the clinical overlap between PD and atypical parkinsonism. The utility of α -Synuclein (α Syn) Seed Amplification Assay (SAA) as a diagnostic indicator for PD has been reported in numerous studies, but never when administered as a validated clinical laboratory test. This study compares results from α Syn-SAA validation testing performed using well-characterized cohorts from two biorepositories to better understand the accuracy of PD clinical diagnosis. Blinded cerebrospinal fluid (CSF) specimens from a repository that included cohorts of subjects clinically diagnosed as PD or healthy controls, both with confirmatory dopamine transporter single-photon emission computed tomography (DAT SPECT) imaging, and blinded CSF specimens from a repository that included cohorts of subjects clinically diagnosed as PD or healthy controls based on clinical diagnosis alone, were tested as part of the validation studies for the diagnostic α Syn-SAA test (SYNTap® Biomarker Test). Measured α Syn-SAA test accuracy was 83.9% using clinical diagnosis as comparator, and 93.6% using clinical diagnosis with confirmatory DAT- SPECT imaging as comparator. The statistically significant discordance between accuracy determinations using specimens classified using different diagnostic inclusion criteria indicates that there is some symbiosis between dopamine-weighted imaging and α Syn-SAA results, both of which are associated with higher accuracy compared with the clinical diagnosis alone.

Keywords α -Synuclein \cdot Seed amplification \cdot Synucleinopathy \cdot Parkinson's disease

Introduction

Approximately 20% of Parkinson's disease (PD) may be misdiagnosed due to the clinical overlap between PD and atypical Parkinsonism [1]. Non-PD Parkinsonism disorders confound the diagnosis of PD and may have different pathological mechanisms (e.g., tauopathy, TDP-43 aggregation, etc.), and management [2]. Dopamine transporter singlephoton emission computed tomography (DAT-SPECT) identifies deficits in presynaptic domine levels and is useful for demonstrating nigrostriatal degeneration occurring in PD and other diagnoses involving dopamine deficit [3, 4]. DAT-SPECT findings reflect dopaminergic function and do

Kendal Jay Jensen Kjensen@amprionme.com not necessarily identify the exact underlying etiology when a deficit is identified. However, when used for the indication of aiding PD diagnosis in cohorts with suspected PD (high pre-test probability), DAT-SPECT strengthens the accuracy of diagnosis [5]. The Parkinson's Progressive Markers Initiative (PPMI) study included DAT-SPECT imaging to enhance the accuracy of diagnosis for PD and control cohorts with the rationale that individuals with "scans without DAT deficit are unlikely to have PD." [5]. An accurate PD biomarker test is therefore expected to have a high correlation to PPMI cohort assignment, since there is high confidence in the accuracy of clinical diagnosis associated with these specimens.

The α -Synuclein (α Syn) Seed Amplification Assay (SAA) is a novel biomarker test that has significant support in the clinical research community. The assay utilizes in vitro propagation of minute amounts of aggregated misfolded α Syn to achieve levels detectable by simple fluorescence measurements of the amyloid-specific dye, Thioflavin T (ThT) [6–9]. α Syn-SAA reproducibility across laboratories

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and methodological variations has been previously demonstrated; however, no study has been performed in a CLIA (Clinical Laboratory Improvement Amendments)/CAP (College of American Pathologists) accredited laboratory, and no study has compared cohorts selected with and without diagnostic imaging using SAA as the comparator [10]. To this end, we compared assay results between two cerebrospinal fluid (CSF) repositories using a CLIA/CAP validated α Syn-SAA, and compared test performance to repository inclusion diagnostic criteria.

Materials and methods

All patient specimens were collected under institutional review board approved protocols and with informed consent.

Study populations

For the first study, clinical data and frozen CSF from 98 participants were obtained from the Parkinson's Disease Biomarkers Program (PDBP) biorepository [11]. PDBP is a consortium in which participants are assessed longitudinally using standardized sample collection protocols and clinical assessments using Movement Disorder Society or UK Brain Bank criteria [12]. We initially identified case samples from subjects with PD (n=41) and then identified control samples age and sex-matched to case samples to the extent possible (n=57).

For the second study, different specimens from different individuals were obtained from the Parkinson's Progression Markers Initiative (PPMI) repository [5, 10]. Unlike the PDBP repository, the PPMI enrollees require an abnormal DAT-SPECT test in addition to clinical symptoms to be included in the PD cohort and a normal DAT-SPECT test to be included in the control cohort. A total of 343 blinded samples from the PPMI biorepository were tested. The 343 samples were composed of 250 specimens from 109 individuals classified as controls, and 93 specimens collected from 55 individuals classified as PD. Specimens from participants who were clinically diagnosed with PD but had a normal DAT-SPECT on visual inspection (Scans Without Evidence of Dopaminergic Deficit; SWEDD) were not included in this study [5, 13]. In most instances where multiple samples were tested for one subject, these were collected from different visits. To appropriately weigh the influence of individual subjects on the statistics, results within the subject were randomly selected through computer-generated modeling to determine the most likely accuracy metrics for reporting. (Refer to Online Resource for additional information.) Fig. 1 depicts the study populations' testing for both cohorts.



Fig. 1 Flow chart of participants included in the study; the truth tables were constructed using α Syn-SAA results (predictive) compared against the repository clinical classifications (presumed true)

SAA procedure

Briefly, a 2.45 mm diameter borosilicate glass bead (Sigmund Lindner GmbH, cat# 55-02450-89RTS), 160 µL of the reaction mixture, and 40µL of test sample were combined in each test well of a 96-well plate, and the plate was sealed with an optical adhesive film (Applied Biosystems, cat# 4311971). The reaction mixture used throughout the validation studies was similar to formulations described in previous studies [10, 14] and was composed of 100 mM PIPES (MilliporeSigma, cat#80635) pH 6.5, 10 µM ThT (MilliporeSigma, cat#T3516-25G), nuclease-free water (Growcells, cat#NUPW-0500), 500 mM NaCl (Lonza, cat#51202), and 0.3 mg/mL recombinant monomeric human αSyn (Amprion, cat#S2020). Three test wells are used for each patient assessment. A BMG LABTECH FLUOStar Ω Microplate Reader (excitation wavelength, 440 nm; emission wavelength, 490 nm) was used to measure ThT fluorescence in relative fluorescence units (RFU). Following a baseline reading, the assay plate was incubated at 37 °C and subjected to cycles of orbital shaking at 800 rpm for 1 min followed by a pause for 29 min. Fluorescence readings were taken once per day for 7 days with the exception of infrequent extended cases

requiring 10 days. Extension occurs when a partial fluorescence signal has evolved near the end of the 7-day read window. Following the final measurement, the maximum relative fluorescence units (RFU) of each well is determined and the median of the 3 wells for each sample is calculated. Samples with median signal values greater than or equal to 25,000 RFU are classified as "Detected" (positive for aggregates of misfolded α Syn), and samples with median signal values < 25,000 RFU are classified as "Not Detected" (negative for aggregates of misfolded α Syn).

Statistics

The positive predictive value, negative predictive value, sensitivity, specificity, accuracy, and error rate (false results divided by the total number of results) are reported. 95% confidence intervals were calculated using the binomial confidence interval calculator in https://statpages.info/confi

nt.html. Graphs and tables were generated using Microsoft Excel. Statistics were performed using R version 4.1.1 with statistical significance set to 0.05.

Results

We first looked at potential demographic differences between PPMI and PDBP repositories and found no statistical differences in age, race, or ethnicity apart from race and gender for the control cohort comparison (Table 1). To verify the PDBP and PPMI cohorts' comparability in terms of symptomology, United Parkinson's Disease Rating Scale (UPDRS) data were analyzed; there were no statistical differences between repositories (Table 1, Fig. 2).

An important difference between these repositories is the inclusion criteria, which includes DAT-SPECT in the PPMI cohort. Considering repository diagnosis as the comparator,

 Table 1
 Summary of Cohort Parameters. UPDRS comparisons were performed using robust T tests (Yuen), age comparisons were performed using the Kruskal–Wallis Rank Sum Test, and Race Ethnicity Sex used Pearson's Chi-squared Tests

| Parameter | Subject group | | | | | | | <i>p</i> values from repository compari- sons | |
|------------------|-------------------|-------------|-------------|--------------|------------|-------------|-------|---|--|
| | PPMI reposit | ory | | PDBP reposit | tory | | | | |
| | Case | Control | Total | Case | Control | Total | Case | Control | |
| Total UPDRS scor | res non-parametri | c summary | | | | | | | |
| Median | 50 | 4 | _ | 48 | 5 | _ | 0.727 | 0.371 | |
| IQR | 18.5 | 4 | _ | 22 | 8 | _ | | | |
| Min | 14 | 0 | _ | 17 | 0 | _ | | | |
| Max | 72 | 25 | _ | 104 | 43 | _ | | | |
| Demographics cor | nparison | | | | | | | | |
| Age | | | | | | | | | |
| Mean | 64.0 | 60.1 | 61.4 | 64.3 | 61.4 | 62.6 | 0.932 | 0.882 | |
| Median | 64.0 | 64.0 | 64.0 | 63.6 | 64.0 | 63.6 | | | |
| SD | 9.0 | 11.8 | 11.1 | 8.0 | 10.0 | 9.3 | | | |
| Min | 34.2 | 31.0 | 31.0 | 48.4 | 39.4 | 39.4 | | | |
| Max | 77.5 | 85.1 | 85.1 | 81.0 | 84.4 | 84.4 | | | |
| < 60 | 16 (29.1%) | 53 (48.6%) | 69 (42.1%) | 12 (29.3%) | 27 (47.4%) | 39 (39.8%) | | | |
| >60 | 39 (70.9%) | 56 (51.4%) | 95 (57.9%) | 29 (70.7%) | 30 (52.6%) | 59 (60.2%) | | | |
| Race | | | | | | | | | |
| Caucasian | 52 (94.5%) | 99 (90.8%) | 151 (92.1%) | 37 (90.2%) | 47 (82.5%) | 84 (85.7%) | 0.395 | 0.041 | |
| Black | 1 (1.8%) | 7 (6.4%) | 8 (4.9%) | 3 (7.3%) | 10 (17.5%) | 13 (13.3%) | | | |
| Other | 2 (3.6%) | 3 (2.8%) | 5 (3.0%) | 1 (2.4%) | 0 (0.0%) | 1 (1.0%) | | | |
| Ethnicity | | | | | | | | | |
| Hispanic | 2 (3.6%) | 2 (1.8%) | 4 (2.4%) | 1 (1.6%) | 2 (3.5%) | 3 (2.5%) | 1.000 | 0.879 | |
| Not Hispanic | 53 (96.4%) | 107 (98.2%) | 160 (97.6%) | 59 (96.7%) | 54 (94.7%) | 113 (95.8%) | | | |
| Unknown | 0 (0.0%) | 0 (0.0%) | 0 (0.0%) | 1 (1.6%) | 1 (1.8%) | 2 (1.7%) | | | |
| Sex | | | | | | | | | |
| Male | 38 (69.1%) | 68 (62.4%) | 106 (64.6%) | 23 (56.1%) | 24 (42.1%) | 47 (48.0%) | 0.274 | 0.020 | |
| Female | 17 (30.9%) | 41 (37.6%) | 58 (35.4%) | 18 (43.9%) | 33 (57.9%) | 51 (52.0%) | | | |



Fig.2 Box and whisker plots of the total UPDRS Scores broken down by repository and case/control cohorts. The median UPDRS scores for the PDBP and the PPMI control and case subjects were 4 and 5 and 48 and 50, respectively. p values from robust T tests (Yuen) were 0.37 and 0.73, respectively, when comparing control and case scores between repositories

 α Syn-SAA accuracy was higher in the PPMI cohort (93.6%) than in the PDBP cohort (83.9%) (Table 2). It is important to note that the error rate for SAA using the PPMI cohort was 6.4% compared to 16.1% in the PDBP cohort (*p* 0.045) (Table 2). For information, additional validation results, including analytical validation and specimen stability studies, are shown in the Online Resource.

Discussion

The α Syn-SAA results reveal discordance in the accuracy of clinical diagnosis between two repositories of samples collected from donors diagnosed with PD based on different criteria, namely clinical diagnosis with and without ancillary imaging/enrichment by DAT SPECT. Discordance was anticipated since clinical diagnosis of PD based on clinical evaluation alone carries a significant misdiagnosis rate of ~ 20% [1, 15] and is known to be enhanced with a demonstration of the presence or absence of degenerative findings by DAT-SPECT [16].

Because of the relation between higher α Syn-SAA accuracy and enrichment by DAT-SPECT in the inclusion

criterion, we endorse that diagnoses associated with specimens obtained from the PPMI are close to clinical truth (precision between Parkinson diagnostic rate and Parkinson pathobiology). Enrichment by aSyn-SAA alone might reach similar or higher PD diagnostic accuracy than clinical diagnosis enhanced by DAT-SPECT, since αSyn-SAA reaches high accuracy in the PPMI cohort (~94%) and DAT-SPECT enrichment is not 100% diagnostic [17]. Moreover, α Syn-SAA has been shown to be more sensitive in borderline PD cases of the PPMI cohort [14]. Dopaminergic depletion is hypothesized to occur later in the disease process than α Syn misfolding, so it is possible that the positive α Syn-SAA samples with normal DAT-SPECT are at earlier stages of the disease. In theory, the provider may find utility in either of these methods depending on the clinical presentation since both DAT-SPECT and αSyn-SAA studies have shown utility in refining PD etiology by uncovering information from two discrete areas in the PD landscape.

These hypotheses were confirmed in a recently published research study using > 1100 PPMI CSF specimens and representing the largest analysis to date of aSyn-SAA [18]. Similar to the results in the present study using PPMI specimens, this study showed sensitivity for sporadic PD of 93.3%, and specificity for healthy controls of 96.3%. The larger PPMI study also included specimens from SWEDD, prodromal, and non-manifesting genetic carriers. Sensitivity for prodromal PD (hyposmia or REM behavior sleep disorder) was 86%. Importantly, there was evidence that abnormal α Syn aggregation detectable by α Syn-SAA occurs before other detectable clinical or biomarker changes, including DAT-SPECT. In addition, aSyn-SAA provided information about molecular heterogeneity, with α Syn-SAA positivity lower in LRRK2 PD (67.5%) and higher in GBA PD (95.9%) compared with sporadic PD. Overall, the authors conclude that α Syn-SAA classifies people with PD with high accuracy and can play a crucial role to identify pathologically defined subgroups of people with PD and to establish a biomarkerdriven definition of the disease. aSyn-SAA technology has been extensively proven in the research environment, and the test has been officially validated per CLIA and CAP guidelines allowing a significant advancement in diagnostic

| Table 2 αSyn-SAA test accuracy summary | Metric | PDBP repository samples # con- trols—57, # PD—41 | PPMI repository samples # con- trols—109, # PD—55 | |
|--|-----------------------|---|--|--|
| | Classification method | Clinical diagnosis w/out DAT SPECT | Clinical diagnosis with DAT SPECT | |
| | Accuracy | 85.7% (95% CI=0.77-0.92) | 93.9% (CI=0.89-0.97) | |
| | Sensitivity | 80.5% (95% CI=0.65-0.91) | 87.3% (CI=0.76-0.95) | |
| | Specificity | 89.5% (95% CI=0.78-0.96) | 97.2% (CI=0.92-0.99) | |
| | Error rate | 14.3% (CI=0.080-0.228) | 6.1% (CI=0.030-0.109) | |
| | PPV | 84.6% (95% CI=0.69-0.94) | 94.1% (CI=0.84–0.99) | |
| | NPV | 86.4% (95% CI=0.75-0.94) | 93.8% (CI=0.88-0.98) | |

biochemistry that has the capacity to change the diagnostic approach to neurodegenerative diseases. Accurate "rule-in" and "rule-out" of synucleinopathies with a biochemical test promises utility since diagnosis of synucleinopathies can otherwise consume long periods of time and potentially involve unnecessary drug challenges. This validation also serves as a precedent for a new class of clinical testing using protein aggregation science in a CLIA-certified highcomplexity laboratory. Such a test will also aid in clinical research trials requiring a validated test platform for subject screening or monitoring.

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Data availability statement The data that support the findings of this study are available from the corresponding author upon reasonable request.

Declarations

Conflicts of interest Dr. Jensen consults for Prometheus Laboratories, Sharp Healthcare, and Adaptive Biotechnologies, and holds no patent rights and stocks with these entities. Dr. Concha is an inventor on several patents related to SAA technology. All authors are associated with Amprion Inc., a biotech company focused on the development and commercial utilization of SAA technology.

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