

Black Men's Brain Health Study (BMBHS)

BIOSPECIMEN COLLECTION & PROCESSING

Overview

- 1. Specimen uniformity and quality
- 2. Site Equipment
- 3. Procedures
 - Kit Contents and Ordering
 - Sample Labelling
 - Sample Collection and Processing
 - Shipping Samples
 - Non-Conformance
- 4. Contact Information

Specimen Uniformity and Quality

GENERAL REMINDERS

Specimen Standardization and Quality

Most biomarkers are sensitive to *time* and *temperature*

- Standardization of processing across sites is key
- Specimens must be processed within 2 hours of collection
- Reference the BioSEND Biomarker Specimen Collection, Processing, and Shipment Manual as needed
- Do not replace or supplement any kit components without first receiving approval from BioSEND/NINDS

Questions? Email biosend@iu.edu

Site Consumables and Equipment

Sites will need to supply the following items:

- Gloves
- Alcohol wipes
- Butterfly needles
- Tourniquet
- Gauze pads
- Bandages
- Microcentrifuge tube rack
- Sharps bin and lid

- Crushed ice
- Pipettes and pipette tips
- Centrifuge capable of maintaining 4°C
- -80°C Freezer
- Dry ice

Procedures

MAINTAINING SPECIMEN UNIFORMITY AND QUALITY

Biospecimen Collection Protocol

Collection Tube	Drawn At	Specimen Type	Aliquot Volume	Total Number of Aliquots	Cryovial Cap Color	Shipping Temperature
2 EDTA (plastic) Blood	Visit 1	Plasma	1.5ml	6	Purple	Frozen
Collection Tubes, 10ml	Visit 1	Buffy Coat	~750ul	2	Clear	Frozen

Kit Contents and Ordering

- All sites will be sent a Supplemental Kit with their first kit shipment
 - Contains extra blood collection tubes and processing supplies
 - May be used to replace items in study visit kits
- Study Visit Kits should be ordered as soon as visits are planned
 - Contains collection and processing supplies for 1 subjects visit
 - Includes barcoded labels
 - The supplies/labels in each study visit kit are intended for that visit only

Kit Contents and Ordering – REDCap Survey

http://kits.iu.edu/biosend/bmbhs

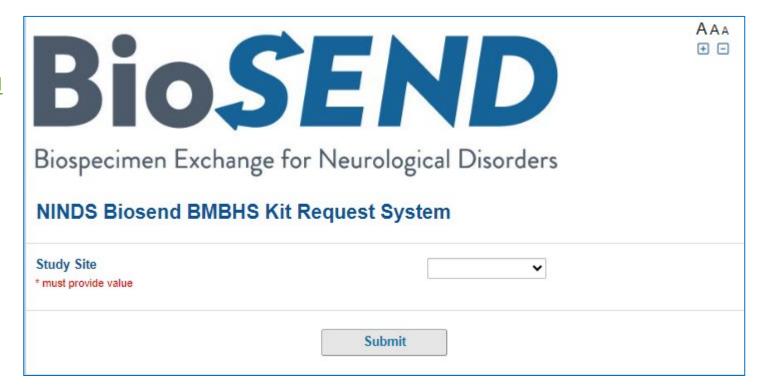
(Boston site will order kits through NCRAD:

https://redcap.uits.iu.edu/surveys/?s=H
X7MARPP8P)

Order kits online through the Kit Request Module for:

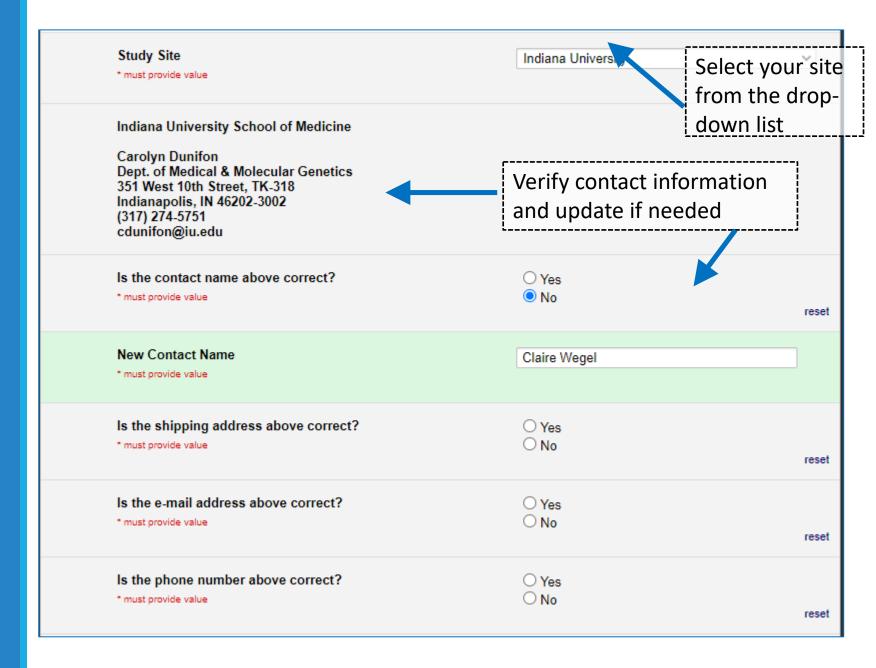
- Blood kits
- Shipping kits
- Extra Supplies

Please provide as much notice as possible when ordering kits and/or supplies.



Kit Contents and Ordering: Confirm Site Info

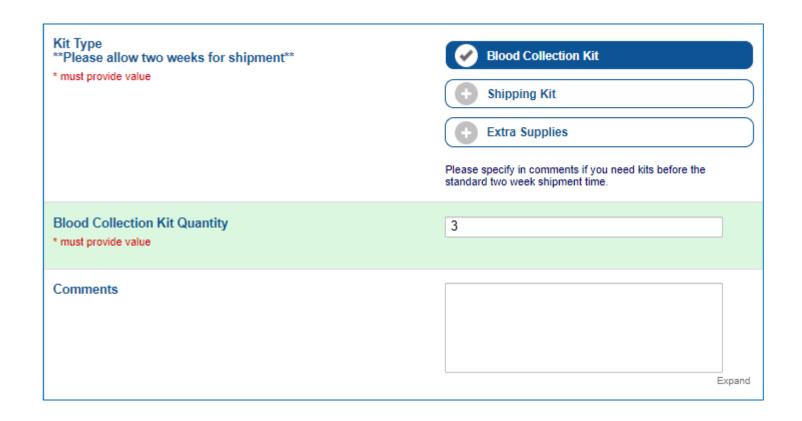
BMBHS Kit Request Module



Kit Contents and Ordering: Kit Types

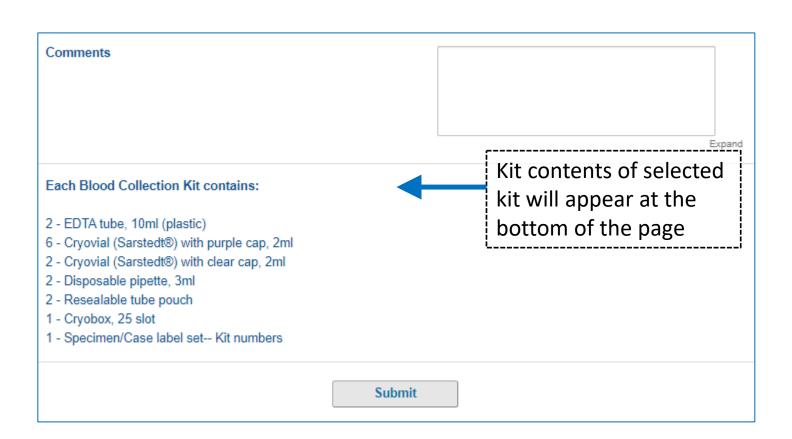
BMBHS Kit Request Module

- Kits are not specific to a subject or time point. After collection, sites will indicate the subject and time point to which BioSEND should link the samples.
- Standard collection kit for plasma and buffy coat contains supplies for one subject-visit.
- Shipping Kits are ordered independently of Blood Kits
- A single Shipping Kit may be used to send one to two cryoboxes



Kit Contents and Ordering: Kit Breakdown

BMBHS Kit Request Module



Kit Contents and Ordering: Kits

Blood Kit (frozen):



Cryobox will be excluded from kits going to Boston since samples are shipping back via NCRAD.

Shipping Kit:



Kit and Supply Ordering

- Click "Submit" to send order to BioSEND; staff will confirm receipt of your order
- Please allow two week turn-around time for kit shipments
- If urgent request needed, please note date needed by in comments and email BioSEND. We cannot guarantee urgent orders, but we will do our best to accommodate.
- BioSEND will send confirmation of shipment and tracking number when supplies ship

Sample Labelling: Example Labels

Labels are provided by Indiana University

- Please check that all samples are properly labelled to ensure correct identification by IU
- If do not have enough labels to complete a visit, please contact IU immediately
- Labelling the tubes during processing prevents sample mix-ups



0005482315 BioSEND

Kit: 302326

PLASMA

Sample Labelling: Label Placement

Please...

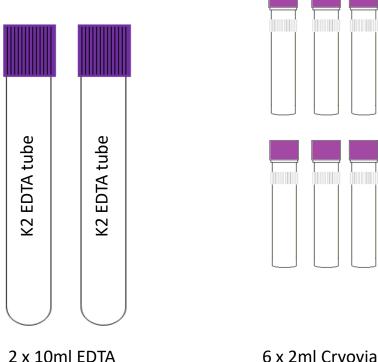
- Label all collection and aliquot tubes before cooling, collecting, processing, or freezing samples
- Label only 1 subject's tubes at a time to avoid mix-ups
- Wrap the label around the tube horizontally label position is important for all tube types
- Make sure the label is completely adhered by rolling between your fingers





Sample Collection and Processing

Supplies provided for the collection of plasma and buffy coat



(plastic) collection

tubes

6 x 2ml Cryovials (purple-capped) for plasma aliquots



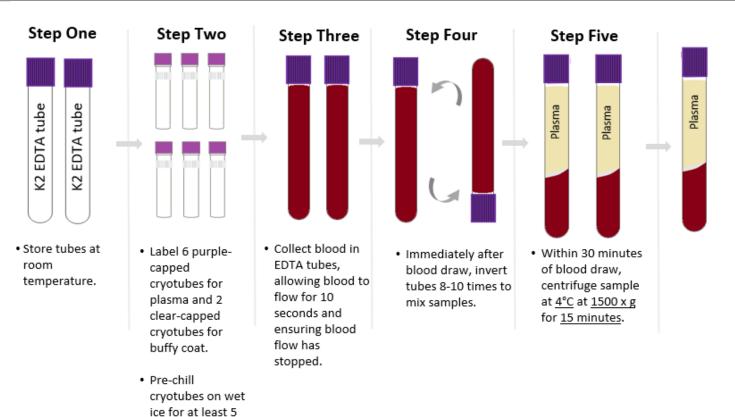
2 x 2ml Cryovials (clear-capped) for buffy coat aliquots

Collection Volumes

Total blood volumes

Sample Type	Amount
Whole Blood for Plasma and Buffy Coat	20 ml

Sample Collection and Processing: Plasma & Buffy Coat



minutes.

Step Six



 Using a clean transfer pipet, aliquot 1.5 ml plasma into each purple cryotube.

Step Seven • Using a clean

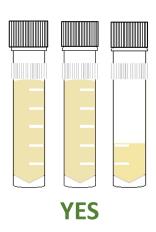


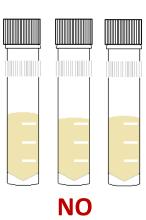
- Using a clean transfer pipet, collect the buffy coat (will have residual plasma and some RBCs included).
- Transfer the buffy coat from each EDTA tube into a clearcapped cryotube.
- Store plasma & buffy coat <u>upright</u> at -80 until shipment.

Sample Collection and Processing: Aliquots

Filling biomarker plasma aliquots:

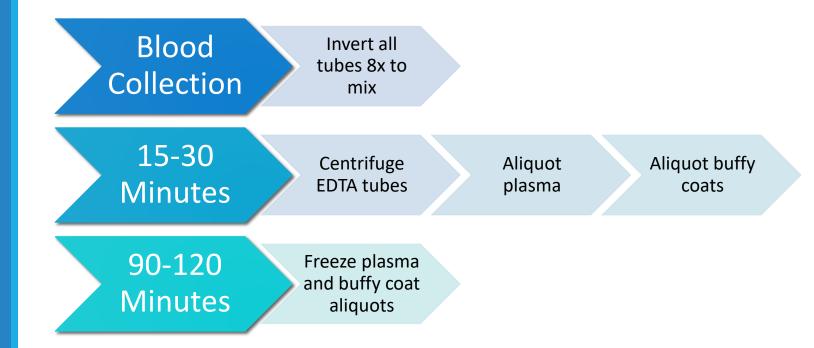
- Fill as many cryovials as possible to 1.5 ml (plasma)
- Over-filled vials may burst in freezer!
- Ship ALL material to IU, even if final vial is less than standard volume





Sample Collection and Processing: Timeline

Timeline for blood processing



Sample Collection and Processing: Issue #1

Troubleshooting Blood Collection

Issue #1: Tube with little/no vacuum

- Always check expiration date on the tube before beginning blood draw and discard expired tubes
- Store tubes at "room temperature" extreme temperature can affect vacuum
- Keep extra vacutainer tubes from supplemental kit nearby during blood draw to replace "bad" tubes
- If this is a frequent occurrence, report tube type and lot number to IU.

Sample Collection and Processing: Issue #2

Troubleshooting Blood Collection

Issue #2: Hemolyzed serum and/or plasma caused by incorrect collection

Cause: Blood Collection Methods	Corrective Action	
Improper venipuncture site	Draw from median cubital, basalic, and cephalic veins from antecubital region of arm	
Prolonged tourniquet use	Tourniquet should be released after no more than 1 min, excessive fist clenching should be avoided	
Not allowing alcohol to dry on skin before venipuncture	Without touching, allow the venipuncture site to air dry	
Use of too large/small bore needle resulting in excess force applied to blood	Avoid using too small/large needle. Needle size dependent on the subject's physical characteristics & amount of blood to be drawn. Most commonly used sizes are 19 – 23.	
Pulling/pushing plunger too fast while drawing/transferring blood	Avoid drawing the syringe plunger too forcefully when collecting blood	
Ensure all blood collection assemblies are fitted securely, to avoid frothing		

For more information, visit: http://www.bd.com/vacutainer/pdfs/techtalk/TechTalk_Jan2004_VS7167.pdf

Sample Collection and Processing: Issue #2 continued

Troubleshooting Blood Collection

Issue #2: Hemolyzed serum and/or plasma caused by incorrect processing

Cause: Sample Processing Methods	Corrective Actions
Vigorous mixing/shaking	Gently invert blood collection tube when mixing additive with specimen, follow guidelines in Biologics Manual regarding number of times to invert each type of tube
Not allowing serum to clot for recommended time	Serum tubes without clot activator should be allowed to clot for 60 min in a vertical position
Exposure to excessive heat or cold	Keep samples at ambient temperature until processing
Prolonged contact of serum/plasma with cells	Do not store uncentrifuged samples beyond recommended time

For more information, visit: http://www.bd.com/vacutainer/pdfs/techtalk/TechTalk_Jan2004_VS7167.pdf

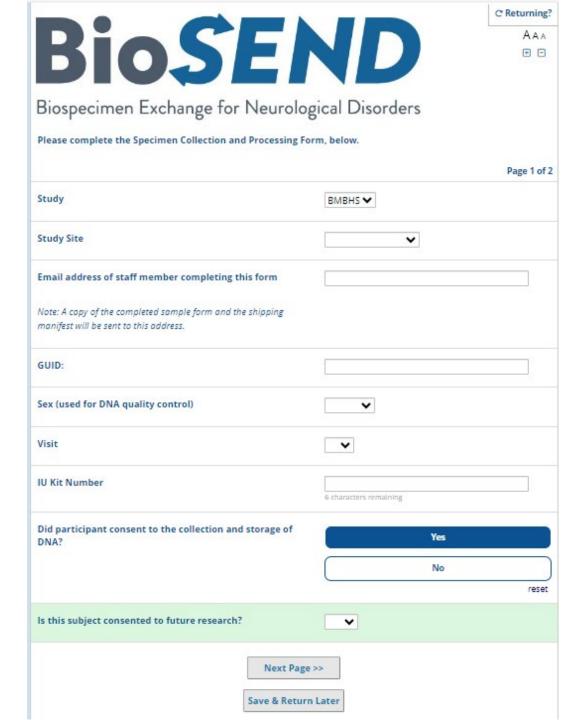
Sample Collection and Processing Form

Specimen Collection and Processing Form

Direct Link to processing form:

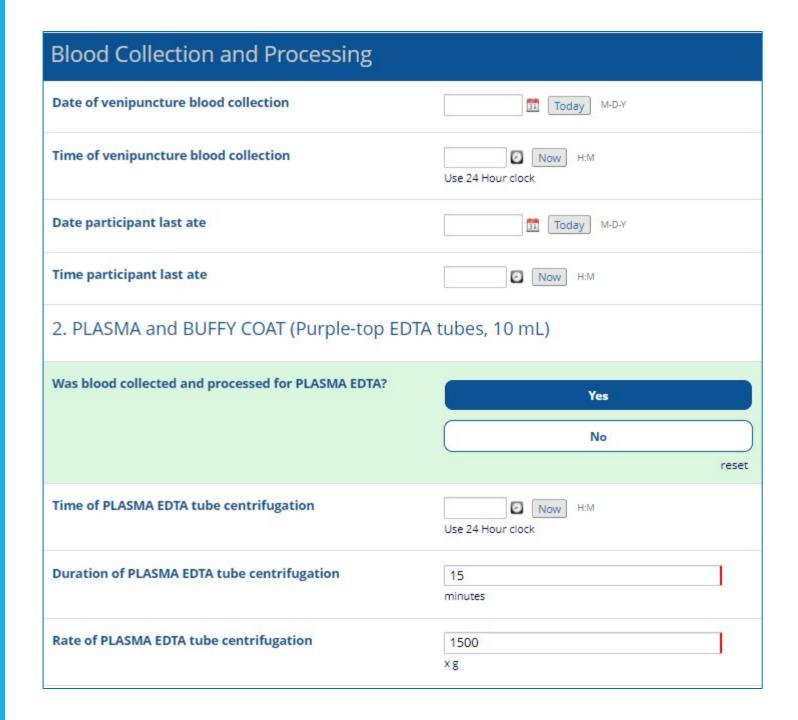
<u>https://kits.iu.edu/biosend/BMBHSS</u> ampleForm

First page captures basic subject and visit information



Sample Collection and Processing Form

Second page captures processing information



Page 1

Sample Collection and

Processing Form

PDF form of responses will be emailed to you. Print a copy of the Frozen Shipping Manifest and include with shipment.

BMBHS Frozen Shipping Manifest

Study

Please verify/update the information below. When you click the "Submit" button below, a PDF copy of the Frozen Shipping Manifest will be emailed to you for Subject [subj_id].

○ BMBHS

Please print a copy of that document and include it in the Kit #[kit num] shipping container.

,	
Study Site:	O Boston University Duke University
GUID:	
Visit:	○ V1
IU Kit Number:	
Date of blood collection:	
PLASMA EDTA	
Number of PLASMA EDTA aliquots shipped:	
Number of BUFFY COAT aliquots shipped:	
Shipping Information - Please complete.	
Frozen shipments should be sent Monday-Wednesday Contact us at biosend@iu.edu if you are unsure wheth	only. Please check for holiday closures prior to shipping. er or not it is safe to ship.

Page 1

Sample Collection and Processing Form

Specimen Collection and Processing Form

A physical copy of the sample form will be included in kits being shipped via NCRAD to Boston site.

First page captures basic subject and visit information

BMBHS Specimen Collection And Processing Form

Please complete the Specimen Collection and Processing Form, below.

Study	O BMBHS
Study Site	O Boston University O Duke University
Email address of staff member completing this form	
Note: A copy of the completed sample form and the shipping manifest will be sent to this address.	
GUID:	
Sex (used for DNA quality control)	O Male O Female O Other
/isit	O V1
U Kit Number	
Did participant consent to the collection and storage of DNA?	O Yes O No
s this subject consented to future research?	O Yes O No

Sample Collection and Processing Form

Second page captures processing information

Blood Collection and Processing		
Date of venipuncture blood collection		
Time of venipuncture blood collection		
	(Use 24 Hour clock)	
Date participant last ate		
Time participant last ate		
		1
2. PLASMA and BUFFY COAT (Purple-top EDTA tubes, 10 mL)		
Nas blood collected and processed for PLASMA EDTA?	O Yes O No	
	O No	
Time of PLASMA EDTA tube centrifugation		
	(Use 24 Hour clock)	
Duration of PLASMA EDTA tube centrifugation		
	(minutes)	
	(minutes)	
Rate of PLASMA EDTA tube centrifugation		
	(x g)	
Temperature of PLASMA EDTA tube centrifugation		
	(degrees Celsius)	
Number of PLASMA EDTA aliquots created for BioSEND		
	(Each aliquot should be 1.5 mL)	
	,	
Number of BUFFY COAT aliquots created for BioSEND		
Time PLASMA EDTA and BUFFY COAT were placed in freezer		
	(Use 24 Hour clock.)	
PLASMA EDTA and BUFFY COAT storage temperature		
EDIN and BOTT CONT Storage temperature		
	(degrees Celsius)	
PLASMA EDTA notes		

Sample Collection and Processing Form

Submission of the Shipping Manifest portion of the form serves as shipment notification to BioSEND

- Must be completed prior to shipment
- If samples are shipped and not received, BioSEND will followup with courier. It is recommended that sites also track shipment to ensure safe delivery

Shipping Frozen Samples: Tips

Packing and Shipping Frozen Samples

- All BMBHS samples ship frozen
- Ship frozen samples on dry ice
- Frozen samples should be shipped only Monday through Wednesday
- Always fill carton to top with dry ice
- Do not pack shipment until the day of pickup



Shipping Samples: Frozen

Each shipping kit contains the supplies to ship two cryoboxes.

- Allows room for dry ice to keep samples frozen in transit
- Minimizes loss in the rare but unfortunate event of courier issues

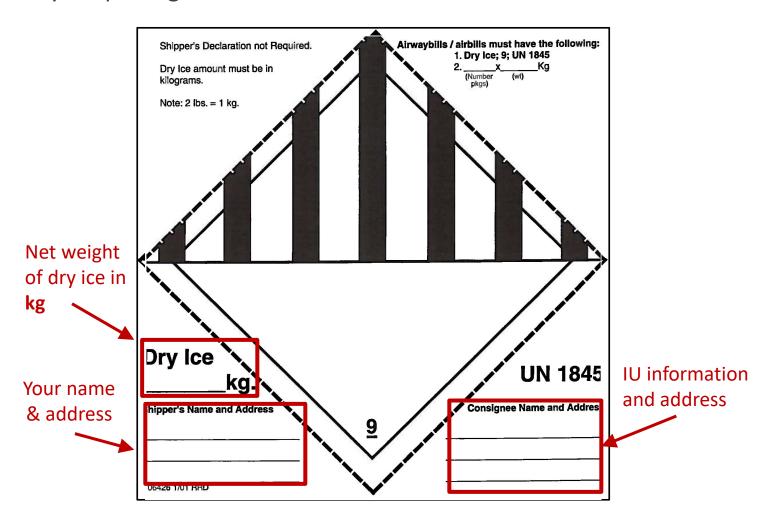




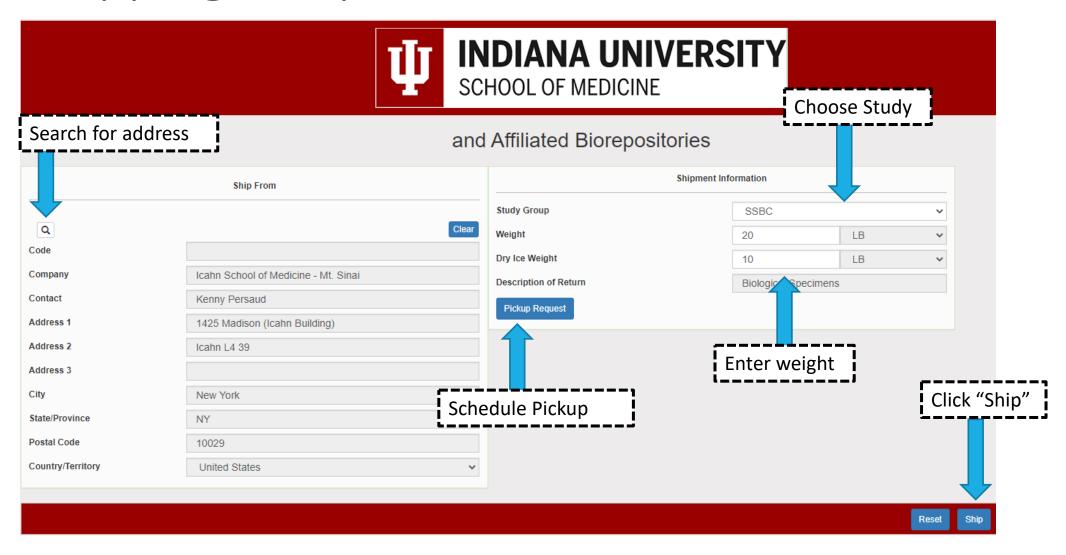
Shipping Samples

Packing and Shipping Frozen Samples

Class 9 Dry Ice Label should not be covered with other stickers and must be completed, or UPS will reject/return your package!



Shipping Samples – UPS: https://kits.iu.edu/UPS



Shipping Samples via UPS

IU UPS ShipExec Shipping Portal

- Print out UPS air waybill
- Ensure all elements
 (barcode, return address, etc.) printed clearly
- Fold and insert UPS air waybill into clear plastic sleeve on package

JOHN SMITH INDIANA UNIVERSITY 410 WEST 10TH STREET INDIANAPOLIS IN 46202 2 LBS

1 OF 1

RS

SHIP TO:

SCHOOL OF MEDICINE 317-278-2694 INDIANA UNIVERSITY TK 217 351 W 10TH ST

INDIANAPOLIS IN 46202



IN 461 9-01



UPS NEXT DAY AIR

TRACKING #: 1Z 976 R8W 84 3985 8595

1



BILLING:

DESC: Biological Specimens RETURN SERVICE

Reference No.1: 4087277

OL 20.03.09 NV45 83.0A 12/



Shipping Samples: Closures

Date	Holiday
January 1	New Year's Day
3 rd Monday in January	Martin Luther King, Jr Day
4 th Monday in May	Memorial Day
June 19	Juneteenth (observed)
July 4	Independence Day (observed)
1 st Monday in September	Labor Day
4 th Thursday in November	Thanksgiving
4 th Friday in November	Friday after Thanksgiving
December 25	Christmas

Please also consider weather when shipping. UPS will post service updates on their webpage. Reach out to BioSEND if you an unsure if it is safe to ship.

Non-Conformance

Non-conformance to standard procedures may reduce the utility of the biospecimens:

- Not processing plasma within 2 hours of collection allows for breakdown of certain proteins and small molecules
- Over/under centrifuging changes plasma composition



Non-Conformance Reporting con't

Most common non-conformance issues:

- Shipment notification not sent
- Samples shipped for weekend/holiday delivery
- Sample form incomplete/inaccurate
- Low volume
- Unlabeled or mislabeled tube(s)
- Sample hemolysis



Non-Conformance and Inventory Reporting

Most common non-conformance issues:

- BioSEND will notify sites directly of any issues upon receipt
- BioSEND will email sites a monthly inventory report of all samples received from that site to date
- If you are experiencing issues, please reach out to us for help! It is much easier to prevent an issue before sample collection & shipment than trying to fix it after the fact

Contacts

Indiana University

General Questions:

biosend@iu.edu

317-278-6158

Request kits:

http://kits.iu.edu/biosend/bmbhs

Boston site will request kits through NCRAD

https://redcap.uits.iu.edu/surveys/?s=HX7MA RPP8P